Positive Control -



Negative Control -



12. SPECIFICITY/INCLUSIVITY

a) 100 % Exclusivity, determined using DNA from 29 non-animal species suitable to occur in the same food products (Table 1).

b) 100 % Inclusivity, determined in 12 food samples obtained from different commercial sources and from External Quality Control (Table 1).

13. SENSITIVITY

The limit of detection (LoD) for SUPREME REAL TIME DETECTION KIT Total Meat is 10 pg of meat DNA per reaction. Analytical sensitivity was determined by a 10-fold dilution series of the target DNA.

The method's detection limit can detect 0.1% of animal DNA in 50 ng of total DNA.

No false negatives were obtained, either in the defined PCR conditions or in the deviations used for robustness, increasing the stringency.

BIOPREMIER

SUPREME

REAL TIME DETECTION KIT

TOTAL MEAT

Ref: BIOPSFS-0067

1. BACKGROUND INFORMATION

Species authenticity is highly relevant to consumers for economic, medical, cultural, and religious reasons. Examples like fraudulent substitution of expensive species with cheaper meats, inclusion of meat in vegetarian products, and allergens in food products highlight the importance of this issue. Traditional protein-based detection methods are unreliable for highly processed and heated products, unlike DNA-based methods. DNA is more stable, and modern PCR techniques enable the identification of fragmented DNA from the species present in a sample, ensuring accurate and highly sensitive detection with a lower detection limit detection. Real time PCR techniques are suitable for amplifying small fragments of DNA high sensitivity and specificity.

2. INTENDED USE

SUPREME REAL TIME DETECTION KIT TOTAL MEAT is designed for the detection of small amounts of meat DNA (down to 10 pg/PCR reaction and 0.1% of meat DNA in 50 ng of total DNA) in a single real-time PCR reaction, after a sample processing extraction step. Therefore, traces of mammalian and avian DNA are detected. The kit allows simultaneous, rapid and sensitive detection of DNA of meat in food and feed samples, including raw ingredients, processed foods, and finished products. Please note that this system does not detect DNA from insects, reptiles or fishes. The kit includes an Internal Control (IC). The IC is used to evaluate PCR inhibitors in the sample or to evaluate problems that occurred during PCR preparation/amplification. The kit includes a Master mix for the target and IC, primers and TaqMan® probes, labelled with non-fluorescent quenchers. The kit contains Uracil-DNA Glycosylase (UDG), preventing DNA contamination with PCR products. The kit includes a positive control, which allows an evaluation of the primers and probes used to detect the targets, and a negative control (nuclease-free water) to confirm the integrity of the kit reagents.

The kit was validated in the instruments ABI PRISM® 7500 Fast. The kit is compatible with all thermocyclers working in **FAM** and **ROX** channels. The detection kit must not be used for diagnostic procedures. For Food use only.

The procedure includes the following main steps:



3. TRADEMARKS AND DISCLAIMER

Registered names, trademarks, etc. used in this document, even when not specifically marked as such, are not to be considered unprotected by law. BPMR kit handbooks and user manuals can be requested from BPMR or your local distributor.

4. LIMITED LICENSE AGREEMENT

Use of this product signifies the agreement of the following terms: The kit must be used solely in accordance with the respective Instructions for Use. BPMR grants no license under any of its intellectual property to use or incorporate the enclosed components of this Kit with any components not included within this kit except as described in the Instructions for Use. This kit and its components are licensed for one-time use and may not be reused, refurbished, or resold.

BPMR – Production and Development, Lda. Amadora INOVA
Rua Henrique de Paiva Couceiro, 10
2700 – 453 Falagueira – Venda Nova Amadora, PORTUGAL
Tel: +351 211 451 410
E-mail: sales.support@biopremier.com
Website: http://www.biopremier.com

BPMR is certified ISO 9001:2015

5. QUALITY CONTROL

In accordance with BPMR's ISO 9001, each lot of the kit is tested against predetermined specifications to ensure consistent product quality.

6. WARNINGS AND PRECAUTIONS

Molecular Biology procedures, such as DNA extractions and PCR amplification, require qualified staff to prevent the risk of erroneous results, especially due to sample contamination or degradation of the nucleic acids contained in the samples. It is strongly recommended to have dedicated areas, materials and equipment for the DNA extraction, preparation of the PCR and post-PCR procedures. Workflow in the laboratory must proceed in a unidirectional manner, beginning in the Extraction Area and moving to the Amplification and Detection Area.

- The user should always pay attention to the following:
- Read all the instructions provided before running the assay.
- Do not mix reagents from different batches.
- Wear proper PPE, including disposable gloves and laboratory coats.
- Store and extract positive material separately from all other reagents.

7. CONTENTS AND STORAGE CONDITIONS

The kit should be stored between -20°C and 5°C and protected from exposure to light. Repeated thawing and freezing (>5) should be avoided, as this may reduce the sensitivity. Considerate to freeze in aliquots to maintain the performance of the assay.

The kit contains reagents for 100 reactions.

CONTENTS		UNITS	COMPOSITION
Master Mix (blue cap)		2 tube (2 x 683µl)	Buffer, dNTPs, Primers and Probes
Enzymes (yellow cap)	•	1 tube (1 x 220µl)	DNA polymerase, UDG and storage buffer
Negative Control (clear cap)	0	1 tube (1 x 130µl)	Nuclease-free water
Positive Control (red cap)		1 tube (1 x 130µl)	Target DNA

8. MATERIAL REQUIRED AND NOT SUPPLIED

- Microcentrifuge
- Laminar Air Flow Cabinets/PCR Cabinets
- Disposable powder-free gloves
- Micropipettes and nuclease-free filter tips
- Real time PCR instrument
- Tubes/Strips/Multiwell plates and accessories specific for each
 Instrument
- DNA extraction kit (example: BIOPEXT-0609)

9. PROCEDURE

9.1 Sample preparation

Depending on the specific characteristics of the sample, procedures like homogenization and grinding may be necessary before DNA extraction.

9.2 DNA extraction

Use a kit or protocol suitable for DNA extraction from food products, such as BIOPREMIER DNA Extraction Kit from Food (ref: BIOPEXT-0609) or similiar. Follow the manufacturer's or authors instructions.

9.3 PCR preparation

A – PCR mix

Always wear gloves for all PCR procedures.

- 1. Thaw the kit solutions. Mix thoroughly (<u>do not vortex the tube</u> <u>Enzymes</u>) and centrifuge briefly
- 2. Prepare the reactions, as described below:

CONTENTS		N° OF SAMPLES		
qPCR reaction		1	10 (10 + 1)	
Master Mix		13 µl	143 µl	
Enzymes	•	2 µl	22 µl	
Total Volume		15 µl	165 µl	

Note: Prepare the PCR reaction for each sample, or in alternative, prepare a Master Mix for the total number of reactions plus 10% (e.g. for 10 samples, prepare a volume for 11). In this case, prepare the Mix in a 1,5mL sterile, nuclease-free tube. Include 2 PCR reactions for the Positive and Negative controls.

- 3. Mix the prepared Mix by inverting the tube and centrifuge briefly
- 4. Dispense 15 μL aliquots of prepared Mix into the plate wells or PCR tube
- For the negative control, pipette 5 µL of Negative Control tube (Clear Cap); Pipette 5 µL of DNA sample per well; and for the positive control, pipette 5 µL of Positive Control tube (Red Cap). Each PCR tube / well should have a final PCR volume of 20 µL
- 6. Centrifuge briefly the plate wells or PCR tubes
- 7. Place the reactions into the Real Time PCR instrument.

B – Program set up

Prepare the Real-Time PCR instrument according to the following temperature/time program:

PHASE	TEMPERATURE	CYCLES	TIME	ACQUISITION
Incubation	37 °C	1	15 min	No
UDG Inactivation	95 °C	1	5 min	No
	94 °C		15 s	No
DNA Amplification	60 °C	45	30 s	Collect Data
	72 °C		15 s	No

Sample Volume: 20 µl Passive reference dye: None Detection format: Hydrolysis Probe

TARGET	CHANNELS		
Detection of meat DNA	FAM Excitation at 465 nm, Emission at 510 nm		
Detection of Internal Control (IC)	ROX Excitation at 533 nm, Emission at 610 nm		

14. DATA ANALYSIS

For analysis of PCR results, select fluorescence display options. Samples with positive Ct-values are considered positive.

Important: Please, also check amplification curves, not only Ct values. Samples should be inspected both in logarithmic and linear scale view and compared with the negative control. Adjust the Threshold, if necessary. Sample results should be assessed after the positive and negative controls have been examined and determined to be valid. If the results of controls are not valid, the sample results cannot be interpreted. Interpretation of PCR-data:

a) Controls

To validate the assay, the controls must have the following results:

	Meat DETECTION FAM	IC DETECTION ROX
Negative Control	Negative	Positive
Positive Control	Positive	Positive

Note that if the controls do not match these results, the experiment must be repeated.

b) Samples

Interpretation of sample results is summarized in the following table:

Meat DETECTION FAM	IC DETECTION ROX	INTERPRETATION	
Positive	Positive/Negative** DNA detected, sample Positive		
Negative	Positive	No DNA detected, sample Negative for meat	
Negative	Negative	Invalid Result*	

*When both Meat and IC detection are Negative, means the presence of PCR inhibitors in the sample. Dilute the sample or perform another DNA extraction.

** High DNA concentration of the target in the sample can lead to a reduced or absent fluorescence signal of the IC.

Table 1: Exclusivity and Inclusivity of the SUPREME REAL TIME DETECTION KIT TOTAL MEAT

Species	SUPREME Kit Result	Result comparison
Tofu VG9	NEG	Matched
Veggie burger VG10	NEG	Matched
100 % vegetable coxinha VG8	NEG	Matched
Soup VG6	NEG	Matched
Vegetarian meatballs C66	NEG	Matched
Potato C20	NEG	Matched
Soy VR20	NEG	Matched
Gadus morhua P6	NEG	Matched
Theragra chalcogramma P20	NEG	Matched
Sparus aurata P28	NEG	Matched
Solea solea P29	NEG	Matched
Xiphias gladius P30	NEG	Matched
Merluccius merluccius P31	NEG	Matched
Pagrus pagrus P34	NEG	Matched
Xiphias gladius P35	NEG	Matched
Salmo salar P36	NEG	Matched
Thunnus spp. P37	NEG	Matched
Almond AL3	NEG	Matched
Nut AL2	NEG	Matched
Spinach VG7	NEG	Matched
Courgette VG5	NEG	Matched
Cabbage VG3	NEG	Matched
Leek VG2	NEG	Matched
Lettuce VG1	NEG	Matched
Sesame AL7	NEG	Matched
Cashew AL6	NEG	Matched
Pecan nut AL5	NEG	Matched
Celery AL4	NEG	Matched
Mustard AL1	NEG	Matched
Equus caballus C5	POS	Matched
Anas platyrhynchos C9	POS	Matched
Meleagris gallopavo C15	POS	Matched
Gallus gallus domesticus C17	POS	Matched
Anser domesticus C26	POS	Matched
Orvctolagus cuniculus C29	POS	Matched
Capra aegagrus hircus (Goat cheese) C33	POS	Matched
Sus scrofa domesticus C56	POS	Matched
Bos taurus (Butter) C59	POS	Matched
Bos taurus (Cow milk) C63	POS	Matched
Ovis aries (Sheep yogurt) C61	POS	Matched
Sus scrofa C65	POS	Matched