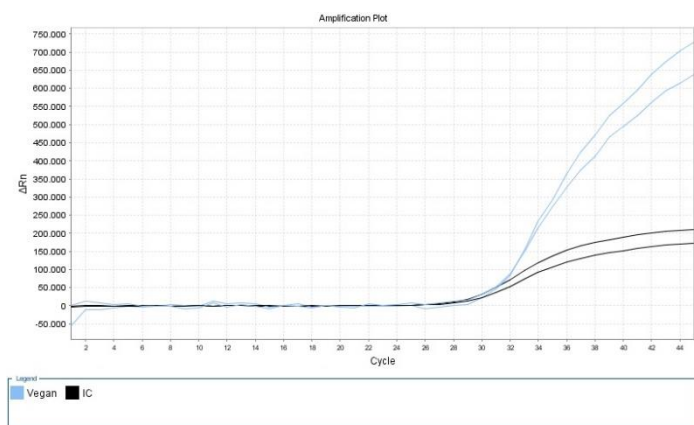
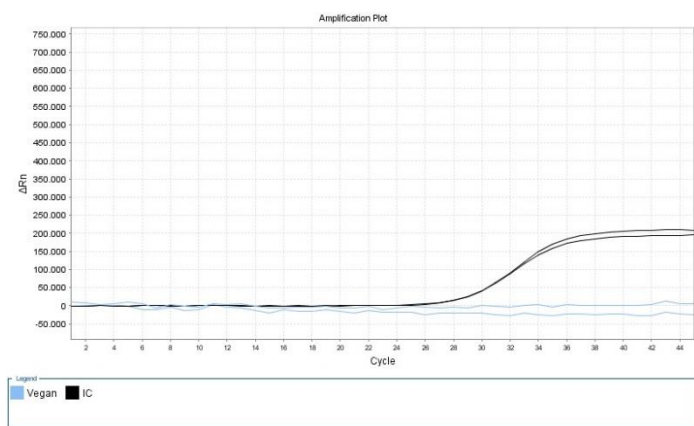


## Positive Control –



## Negative Control –



## 12. SPECIFICITY/INCLUSIVITY

- 100 % Exclusivity, determined using DNA from 19 non-animal species suitable to occur in the same food products (Table 1).
- 100 % Inclusivity, determined in 22 food samples obtained from different commercial sources and from External Quality Control (Table 1).

## 13. SENSITIVITY

The limit of detection (LoD) for SUPREME REAL TIME DETECTION KIT VEGAN is 10 pg of animal DNA per reaction. Analytical sensitivity was determined by a 10-fold dilution series of the target DNA.

The method's detection limit can detect 0.1% of animal DNA in 50 ng of total DNA.

No false negatives were obtained, either in the defined PCR conditions or in the deviations used for robustness, increasing the stringency.



## SUPREME

## REAL TIME DETECTION KIT

## VEGAN

Ref: BIOPSFS-0066

## 1. BACKGROUND INFORMATION

Species authenticity is highly relevant to consumers for economic, medical, cultural, and religious reasons. Examples like fraudulent substitution of expensive species with cheaper meats, inclusion of meat in vegetarian products, and allergens in food products highlight the importance of this issue. With a growing number of individuals adopting vegetarian or vegan lifestyles, it becomes essential to certify the absence of animal-derived ingredients in vegan food to maintain consumer confidence in the supply chain and label claims. Products that advertise being vegan must be checked for this during or after production by food production laboratories or control authorities to protect the consumer from fraud or mislabeling. The, e.g., European V-label defines in its guidelines for unintentional traces of animal residues a limit of less than 0.1% (1 g/kg) of animal ingredients. Traditional protein-based detection methods are unreliable for highly processed and heated products, unlike DNA-based methods. DNA is more stable, and modern PCR techniques enable the identification of fragmented DNA from the species present in a sample, ensuring accurate and highly sensitive detection with a lower detection limit detection. Real time PCR techniques are suitable for amplifying small fragments of DNA high sensitivity and specificity.

## 2. INTENDED USE

SUPREME REAL TIME DETECTION KIT TOTAL VEGAN is designed for the detection of small amounts of animal DNA (down to 10 pg/PCR reaction and 0.1% of animal DNA in 50 ng of total DNA) in a single real-time PCR reaction, after a sample processing extraction step. Therefore, traces of mammalian, avian and fish DNA are detected. The kit allows simultaneous, rapid and sensitive detection of animal DNA in food and feed samples, including raw ingredients, processed foods, and finished products. Please note that this system does not detect DNA from insects, reptiles, shellfish or cartilaginous fish.

The kit includes an Internal Control (IC). The IC is used to evaluate PCR inhibitors in the sample or to evaluate problems that occurred during PCR preparation/amplification. The kit includes a Master mix for the target and IC, primers and TaqMan® probes, labelled with non-fluorescent quenchers. The kit contains Uracil-DNA Glycosylase (UDG), preventing DNA contamination with PCR products. The kit includes a positive control, which allows an evaluation of the primers and probes used to detect the targets, and a negative control (nuclease-free water) to confirm the integrity of the kit reagents.

The kit was validated in the instruments ABI PRISM® 7500 Fast. The kit is compatible with all thermocyclers working in **FAM** and **ROX** channels. The detection kit must not be used for diagnostic procedures. For Food use only. The procedure includes the following main steps:



Sample  
Processing



DNA  
extraction



PCR  
set-up and run



Results and  
Interpretation

## 3. TRADEMARKS AND DISCLAIMER

Registered names, trademarks, etc. used in this document, even when not specifically marked as such, are not to be considered unprotected by law. BPMR kit handbooks and user manuals can be requested from BPMR or your local distributor.

## 4. LIMITED LICENSE AGREEMENT

Use of this product signifies the agreement of the following terms: The kit must be used solely in accordance with the respective Instructions for Use. BPMR grants no license under any of its intellectual property to use or incorporate the enclosed components of this Kit with any components not included within this kit except as described in the Instructions for Use. This kit and its components are licensed for one-time use and may not be reused, refurbished, or resold.



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BPMR is certified ISO 9001:2015

In accordance with BPMP's ISO 9001, each lot of the kit is tested against predetermined specifications to ensure consistent product quality.

Molecular Biology procedures, such as DNA extractions and PCR amplification, require qualified staff to prevent the risk of erroneous results, especially due to sample contamination or degradation of the nucleic acids contained in the samples. It is strongly recommended to have dedicated areas, materials and equipment for the DNA extraction, preparation of the PCR and post-PCR procedures. Workflow in the laboratory must proceed in a unidirectional manner, beginning in the Extraction Area and moving to the Amplification and Detection Area.

- The user should always pay attention to the following:
- Read all the instructions provided before running the assay.
- Do not mix reagents from different batches.
- Wear proper PPE, including disposable gloves and laboratory coats.
- Store and extract positive material separately from all other reagents.

The kit should be stored between -20°C and 5°C and protected from exposure to light. Repeated thawing and freezing (>5) should be avoided, as this may reduce the sensitivity. Considerate to freeze in aliquots to maintain the performance of the assay.

The kit contains reagents for 100 reactions.

## 8. MATERIAL REQUIRED AND NOT SUPPLIED

- Microcentrifuge
- Laminar Air Flow Cabinets/PCR Cabinets
- Disposable powder-free gloves
- Micropipettes and nuclease-free filter tips
- Real time PCR instrument
- Tubes/Strips/Multiwell plates and accessories specific for each Instrument
- DNA extraction kit (example: BIOEXT-0609)

### 9.1 Sample preparation

Depending on the specific characteristics of the sample, procedures like homogenization and grinding may be necessary before DNA extraction.

Use a kit or protocol suitable for DNA extraction from food products, such as BIOPREMIER DNA Extraction Kit from Food (ref: BIOEXT-0609) or similar. Follow the manufacturer's or authors instructions.

### A – PCR mix

Always wear gloves for all PCR procedures.

1. Thaw the kit solutions. Mix thoroughly (do not vortex the tube Enzymes) and centrifuge briefly
2. Prepare the reactions, as described below:

Note: Prepare the PCR reaction for each sample, or in alternative, prepare a Master Mix for the total number of reactions plus 10% (e.g. for 10 samples, prepare a volume for 11). In this case, prepare the Mix in a 1,5mL sterile, nuclease-free tube. Include 2 PCR reactions for the Positive and Negative controls.

3. Mix the prepared Mix by inverting the tube and centrifuge briefly
4. Dispense 15  $\mu$ L aliquots of prepared Mix into the plate wells or PCR tube
5. For the negative control, pipette 5  $\mu$ L of Negative Control tube (Clear Cap); Pipette 5  $\mu$ L of DNA sample per well; and for the positive control, pipette 5  $\mu$ L of Positive Control tube (Red Cap). Each PCR tube / well should have a final PCR volume of 20  $\mu$ L
6. Centrifuge briefly the plate wells or PCR tubes
7. Place the reactions into the Real Time PCR instrument.

Prepare the Real-Time PCR instrument according to the following temperature/time program:

Sample Volume: 20 µl      Detection format: Hydrolysis Probe  
Passive reference dye: None

## 14. DATA ANALYSIS

For analysis of PCR results, select fluorescence display options. Samples with positive Ct-values are considered positive.

**Important:** Please, also check amplification curves, not only Ct values. Samples should be inspected both in logarithmic and linear scale view and compared with the negative control. Adjust the Threshold, if necessary. Sample results should be assessed after the positive and negative controls have been examined and determined to be valid. If the results of controls are not valid, the sample results cannot be interpreted.

Interpretation of PCR-data:

To validate the assay, the controls must have the following results:

Note that if the controls do not match these results, the experiment must be repeated.

Interpretation of sample results is summarized in the following table:

\*When both Animal and IC detection are Negative, means the presence of PCR inhibitors in the sample. Dilute the sample or perform another DNA extraction.

**\*\* High DNA concentration of the target in the sample can lead to a reduced or absent fluorescence signal of the IC.**

Table 1: Exclusivity and Inclusivity of the  
SUPREME REAL TIME DETECTION KIT TOTAL VEGAN

Species	SUPREME Kit Result	Result comparison
Tofu VG9	NEG	Matched
Veggie burger VG10	NEG	Matched
100 % vegetable coxinha VG8	NEG	Matched
Soup VG6	NEG	Matched
Vegetarian meatballs C66	NEG	Matched
Potato C20	NEG	Matched
Soy VR20	NEG	Matched
Almond AL3	NEG	Matched
Nut AL2	NEG	Matched
Spinach VG7	NEG	Matched
Courgette VG5	NEG	Matched
Cabbage VG3	NEG	Matched
Leek VG2	NEG	Matched
Lettuce VG1	NEG	Matched
Sesame AL7	NEG	Matched
Cashew AL6	NEG	Matched
Pecan nut AL5	NEG	Matched
Celery AL4	NEG	Matched
Mustard AL1	NEG	Matched
<i>Equus caballus</i> C5	POS	Matched
<i>Anas platyrhynchos</i> C9	POS	Matched
<i>Meleagris gallopavo</i> C15	POS	Matched
<i>Gallus gallus domesticus</i> C17	POS	Matched
<i>Anser domesticus</i> C26	POS	Matched
<i>Oryctolagus cuniculus</i> C29	POS	Matched
<i>Capra aegagrus hircus</i> (Goat cheese) C33	POS	Matched
<i>Sus scrofa domesticus</i> C56	POS	Matched
<i>Bos taurus</i> (Butter) C59	POS	Matched
<i>Bos taurus</i> (Cow milk) C63	POS	Matched
<i>Ovis aries</i> (Sheep yogurt) C61	POS	Matched
<i>Sus scrofa</i> C65	POS	Matched
<i>Gadus morhua</i> P6	POS	Matched
<i>Theragra chalcogramma</i> P20	POS	Matched
<i>Sparus aurata</i> P28	POS	Matched
<i>Solea solea</i> P29	POS	Matched
<i>Xiphias gladius</i> P30	POS	Matched
<i>Merluccius merluccius</i> P31	POS	Matched
<i>Pagrus pagrus</i> P34	POS	Matched
<i>Xiphias gladius</i> P35	POS	Matched
<i>Salmo salar</i> P36	POS	Matched
<i>Thunnus spp.</i> P37	POS	Matched